Urine

Urine (MSU) for microscopy; culture & sensitivity (MC&S)

The Microbiology department uses boric acid containers for all routine urines for microscopy and bacteriological culture. Two sizes are available holding 20 or 5 mL. Both have a small amount of white powder and have a red top. **Please note that these are NOT suitable for Chemistry investigations**

The methods of MSU collection are as follows:

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Prior to collection the genital area should be cleaned with tap water. Antiseptics should not be used. If the area is soiled; use soap and water and rinse thoroughly.

**Men**: Retract prepuce. Wash the glans penis.

**Women**: Clean the vulva; first the outer labia; then the inner (clean from front to back). Separate the labia while the specimen is passed. Discard the initial part of the urine sample. Collect middle portion of the stream into a clean (preferable sterile) vessel. Then pour appropriate volume into the boric acid container.

**Urine containers for microbiological analysis.**

The Boric acid pots are only suitable for microbiological analysis; and are designed to prevent bacterial growth and preserve cellular constituents during transit.

Collect MSU following the usual procedure.
Do not discard the white powder.
Fill boric acid pot to the line marked and mix well. This gives the correct concentration of preservative.

Despatch specimen to laboratory as soon as possible.

- Boric acid preserved urine specimens may be stored at room temperature prior to processing.
- They do not need to be placed in a refrigerator.
- Please note; do not use boric acid containers for purposes other than microbiological analysis of urine.
The Boric acid pots are **not suitable** for

- Dipstick tests on urine
- Urine investigation for TB
- Urine for pregnancy tests

A limited number of the white topped standard 60ml pots are available for other microbiological specimens (e.g. pus; fluid; sputum); but these will not be issued for microbiology urine samples.

**Urine for Schistosomiasis/Bilharziasis (3 hour collection)**

Void urine at 11am and discard. Thereafter collect all urine passed until 2 p.m. Empty bladder at 2p.m. and add to collection. Send to lab. specifically requesting microscopy for schistosomiasis. (Only useful for *S. haematobium*). Place sample into a plain container (no preservative); available from Microbiology. Alternatively; a terminal urine sample may be sent. The lab should receive the specimen on the day of collection; or if impossible it should be kept cool and dark and delivered the next day at the latest.

**Catheter Urine**

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Specimens should be obtained by aspiration with a sterile needle and syringe from the self sealing sleeve in the drainage tube. If necessary the tube should be clamped below the level of the sleeve to allow urine to collect. Clean sleeve with spirit then insert needle at 45 degree angle (never collect samples from the drainage bag). Place sample in a boric acid container.

**Urine for pneumococcal/legionella antigen**

Send fresh urine specimen to laboratory for referral.